8. A method of producing the bacterial cellulose of claim 1, which comprises culturing cellulose-producing bacteria which produce the bacterial cellulose extracellularly in a culture medium containing a cell division inhibitor, and recovering the bacterial cellulose produced in the culture medium.

9. The method of claim 8, wherein the cell division inhibitor is selected from the group consisting of chloramphenicol, a protein synthesis inhibitor, an organic compound having β-lactamase inhibiting ability, nalidixic acid, promidic acid, pipemidic acid, oxolinaic acid, ofloxacin and enoxacin.

10. The method of claim 9, wherein the protein synthesis inhibitor is selected from the group consisting of tetracycline, puromycin and erythromycin.

11. The method of claim 9, wherein the organic compound having β-lactamase inhibiting ability is thienamycin.

12. The method of claim 8, wherein the concentration of the cell division inhibitor in the culture medium is 0.01 to 5 mM.

13. The method of claim 8, wherein the bacteria are Acetobacter.

14. The method of claim 8, wherein the bacteria are Acetobacter pasteurianus FERM BP-4176.

15. (Amended) The bacterial cellulose of claim 1, wherein the microfibrils have a width of 430 to 1000 nm.

16. (Amended) The bacterial cellulose of claim 1, wherein the microfibrils have a width of 590 to 1000 nm.

17. (Amended) The bacterial cellulose of claim 1, wherein the microfibrils have a Young's modulus of about 13 to 20 GPa.

18. (Amended) The bacterial cellulose of claim 1, wherein the microfibrils have a Young's modulus of about 16 to 20 Gpa.

19. (Amended) The bacterial cellulose of claim 1, wherein the microfibrils have a width of 340 to 1000 nm.

20. (Amended) The bacterial cellulose of claim 1, wherein the microfibrils have a width of 340 to 700 nm.

21. (Amended) The bacterial cellulose of claim 1, wherein the microfibrils have a width of 340 to 600 nm.--

Please add the following claims.

--22. (New) The bacterial cellulose of claim 1, wherein the microfibrils have a thickness of 2.5, 3, 6, or 9 nm.

23. (New) The bacterial cellulose of claim 1, wherein the ratio of the major axis to the minor axis of the microfibrils is about 28:1.0 to 1000:1.0

24. (New) The bacterial cellulose of claim 1, wherein the ratio of the major axis to the minor axis of the microfibrils is about 28:1.0 to 280:1.0.

25. (New) A bacterial cellulose produced by Acetobacter pasteurianus FERM BP-4176 which comprises microfibrils having a thickness of 1 to 9 nm and a width of 250 to 1000 nm.

26. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a width of 250 to 700 nm.

27. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a width of 250 to 600 nm.

28. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a width of 430 to 1000 nm.

29. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a width of 590 to 1000 nm.

30. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a width of 340 to 1000 nm.

31. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a width of 340 to 700 nm.

32. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a width of 340 to 600 nm.

33. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a Young's modulus of about 13 to 20 GPa.

34. (New) The bacterial cellulose of claim 25, wherein the microfibrils have a Young's modulus of about 16 to 20 Gpa.

35. (New) The bacterial cellulose of claim 25, wherein the ratio of the major axis to the minor axis of the microfibrils is about 28:1.0 to 1000:1.0.

36. (New) The bacterial cellulose of claim 25, wherein the ratio of the major axis to the minor axis of the microfibrils is about 28:1.0 to 280:1.0.

37. (New) The bacterial cellulose of claim 25, wherein the microfibrils are ribbon-shaped.

38. (New) A method of producing the bacterial cellulose of claim 25, which comprises culturing cellulose-producing bacteria which produce the bacterial cellulose extracellularly in a culture medium containing a cell division inhibitor, and recovering the bacterial cellulose produced in the culture medium.

39. (New) The method of claim 38, wherein the cell division inhibitor is selected from the group consisting of chloramphenicol, a protein synthesis inhibitor, an organic